

7th Edition, revised in April, 2017

(FOR RESEARCH USE ONLY. DO NOT USE IT IN CLINICAL DIAGNOSIS !)

Ochratoxin A(OTA) ELISA Kit

Catalog No: E-TO-E001

96T

This manual must be read attentively and completely before using this product.

If you have any problems, please contact our Technical Service Center for help.

Phone: 240-252-7368(USA)240-252-7376(USA)

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Website: www.elabscience.com

Please kindly provide us the lot number(on the outside of the box) of the kit for more efficient service.

Test principle

This kit uses Indirect-Competitive-ELISA as the method. It can detect OchratoxinA (OTA) in samples, such as grain (rice, peanut, soybean, etc.) and feed. This kit is composed of Micro ELISA Plate, HRP conjugate, antibody, standard and other supplementary reagents. The micro-plate provided in this kit has been pre-coated with OTA. During the reaction, OTA in the samples or standard competes with OTA on the solid phase supporter for sites of OTA antibody. Then Horseradish Peroxidase (HRP) conjugate is added to each micro plate well, and TMB substrate is for color development. There is a negative correlation between the OD value of samples and the concentration of OTA. The concentration of OTA in the samples can be calculated by comparing the OD of the samples to the standard curve.

Technical indicator

Sensitivity: 1 ppb (ng/mL)

Reaction mode: 37°C, 30min ~ 30min ~ 15min

Detection limit: Grain/Feed --- 10 ppb

Cross-reactivity: Ochratoxin A (OTA) --- 100%

Sample recovery rate: Grain/Feed --- 85% ± 15%

Kits components

Item	Specifications
Micro ELISA Plate	96 wells
Standard Liquid (black cap)	1 mL each (0 ppb, 1 ppb, 3 ppb, 9 ppb, 27 ppb, 81 ppb)
HRP Conjugate (red cap)	5.5 mL
Antibody Working Solution (blue cap)	5.5 mL
Substrate Reagent A (white cap)	6 mL
Substrate Reagent B (black cap)	6 mL
Stop Solution (yellow cap)	6 mL
20× Concentrated Wash Buffer (white cap)	40 mL
Product Description	1 copy

Other supplies required

Instrument:Micro-plate reader, Printer, Homogenizer, Nitrogen Evaporators, Oscillators, Centrifuge, Graduated pipette, Balance (sensitivity 0.01g).

High-precision transferpeltor:Single channel (20-200 μ L, 100-1000 μ L), Multichannel (300 μ L).

Reagents:Methanol, NaHCO₃.

Experimental preparation

Bring all reagents and samples to room temperature before use.

Open the micro-plate reader in advance, preheat the instrument, and set the testing parameters.

1 Sample pretreatment Notice: experimental apparatus should be clean, and use disposable pipette tips to avoid cross-contamination during the experiment.

2 Solution preparation

Solution 1:70%Methanol. Methanol (volume): Deionized water (volume) =7:3

Solution 2: 0.1M NaHCO₃ Solution

Dissolve 4.2g of solid NaHCO₃ with 500 mL of deionized water.

3 Sample pretreatment procedure

Pretreatment ofGrain(rice, corn, millet, etc.)and feed:

- (1) Weigh 2g of crushed homogenate into 50 mL EP tube, add 10 mLof 70% methanol(Solution 1), oscillate for 5min, centrifuge at 4000r/min for 10 min at room temperature;
- (2) Take the supernatant, add 1mLof0.1M NaHCO₃ Solution(Solution 2), mix;
- (3) Take 50 μ L for detection and analysis.

Note: Sample dilution factor: 10, minimum detection dose: 10ppb

Assay procedure

Centrifuge the sample again after thawing before the assay. Bring all reagents to room temperature for 30 min before use. **All the reagents should be mixed thoroughly by gently swirling before pipetting. Avoid foaming.**Dilute 40 mL of concentrated wash buffer into 800mL of wash buffer with deionized or distilled water.

1. **Number:** number the sample and standard in order (multiplewells), and keep a record of standard wells and sample wells.
2. **Add sample:** add 50 μ L of Standard or Sample per well, then add 50 μ L of antibody working solution, cover the plate with sealer we provided, oscillate for 5s gently to mix thoroughly, incubate for 30min at 37°C.
3. **Wash:** uncover the sealer carefully, remove the liquid in each well. Immediately add 250 μ L of wash working buffer to each well and wash. Repeat wash procedure for 5 times, 30s intervals/time. Invert the plate and pat it against thick clean absorbent paper (If bubbles exist in the wells, clean tips can be used to prick them).
4. **HRP Conjugate:** add 100 μ L of HRP conjugate to each well, incubate for 30min at 37°C.

5. **Wash:** repeat step 3.
6. **Color Development:** add 50 μ L of substrate solution A to each well, and then add 50 μ L of substrate solution B. Gently oscillate for 5s to mix thoroughly. Incubate shading light for 15min at 37 $^{\circ}$ C. (If the blue color is too shallow, can extend the incubation time properly).
7. **Stop reaction:** add 50 μ L of stop solution to each well, oscillate gently to mix thoroughly.
8. **OD Measurement:** determine the optical density (OD value) of each well at 450 nm with a micro-plate reader (the 450/630 nm double wavelength is recommended). This step should be finished in 10min after stop reaction.

Result analysis

1. **Absorbance (%) = $A/A_0 \times 100\%$**

A: Average absorbance of standard or sample

A_0 : Average absorbance of 0ppb Standard

2. **Drawing and calculation of standard curve**

Create a standard curve by plotting the absorbance percentage of each standard on the y-axis against the log concentration on the x-axis to draw a semi-logarithmic plot. Add average absorbance value of sample to standard curve to get corresponding concentration. If samples have been diluted, the concentration calculated from the standard curve must be multiplied by the dilution factor.

For this kit, it is more convenient to use professional analysis software for accurate and fast analysis on a large number of samples.

Notes

1. Overall OD value will be lower when reagents is not brought to room temperature before use or room temperature is below 25 $^{\circ}$ C.
2. During the washing procedure, if the wells turn dry, it will lead to bad linear standard curve and poor repeatability, move on to the next step immediately after wash.
3. Mix thoroughly and wash the plate completely. The consistency of wash procedure can strongly affect the reproducibility of this ELISA kit.
4. Micro ELISA plate should be covered by plate sealer. Avoid the reagents to strong light.
5. Do not use expired kit and reagents of different batches.
6. TMB should be abandoned if it turns color. When OD value of standard (concentration: 0) < 0.5 unit ($A_{450nm} < 0.5$), it indicates reagent is deteriorated.
7. Stop solution is caustic, avoid contact with skin and eyes.

Storage and valid period

Storage: Store at 2-8 $^{\circ}$ C. Avoid freeze / thaw cycles.

Valid Period: 1 year, production date is on the packing box.

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